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Fructose: Metabolic signal and modern hazard

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Abstract

There is much interest in the role of sweeteners like table sugar (sucrose) and high-fructose corn syrup in obesity and metabolic disease. Both sweeteners consist of glucose and fructose, two six-carbon isomeric sugars, that account for 15% of the Western diet. While glucose ingestion may promote obesity through its effects to stimulate insulin secretion, fructose has unique metabolic effects that promote triglyceride synthesis and fat accumulation. These effects arise from fructose's well known role as a signal of metabolic plenty. Under modern conditions of overnutrition, chronic excess fructose drives features of metabolic syndrome. Emerging evidence further links fructose to cancer and dementia. Here we review the biochemical, molecular, and

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physiological distinctions between fructose and glucose, as well as the endogenous fructose pathway that makes fructose from glucose. Through this review, we highlight the role of fructose not only as a caloric source, but also as a regulator of metabolic health and disease.

Intake of added sugars, such as table sugar (sucrose) and high fructose-corn syrup (HFCS), have increased markedly in the last few centuries in parallel with the rise in obesity and diabetes, with the strongest association being with sugar-sweetened beverages (SSBs)¹. The World Health Organization recommends a total daily energy intake from 'free' sugars of <10%². In the United States and many high-income countries, SSB intake is dropping but overall free sugar consumption still exceeds 10%. In low- and middle-income countries, SSB and overall free sugar consumption continue to rise.^{3,4}

One hypothesis for how added sugars might cause obesity is that the sweet and highly palatable characteristics of sugar encourage excessive caloric intake that leads to positive energy balance⁵. Another hypothesis is that the glucose component in sugar results in chronic stimulation of insulin that favors fat storage⁵. While both hypotheses carry merit, this review focuses on the metabolic effects of fructose, the other sugar present in sucrose and HFCS. Fructose has also been implicated in driving metabolic syndrome⁶, and while a large exposure comes from added sugars, fructose can also be generated from glucose within the body. Despite the evidence for dietary fructose, and the potential importance of endogenous fructose production in metabolic disease, the role of fructose is often less emphasized than the energy-balance or glucose-insulin models in reviews discussing the pathogenesis of obesity and metabolic disease⁵.

The physiological effects of fructose metabolism are distinct from glucose (Table 1). Fructose catabolism bypasses the regulated steps of glycolysis, with high fructose doses triggering ATP depletion, lactate and glycerate production, and fat synthesis⁷. The metabolic effects of fructose appear to serve as a physiological signal to trigger storage of carbohydrate as fat, to enable survival during future times of limited food availability⁸. While these effects were evolutionarily beneficial, the excessive intake of fructose today makes it a hazardous nutrient that contributes to obesity and metabolic disease.

DIFFERENCES IN GLUCOSE AND FRUCTOSE METABOLISM

Glucose Metabolism.

Starches are polysaccharides composed of long chains of glucose molecules, serving as the primary storage form of carbohydrate in plants (like grains, potatoes, and legumes) and the major source of dietary calories worldwide. Starch is broken down into glucose in the intestinal lumen. Some of the glucose is metabolized as it passes through the intestinal wall and liver with the rest entering the systemic circulation where it is the most abundant circulating carbon source (5 – 10 mM). The rise in blood glucose stimulates a release in insulin that is further enhanced by the gut stimulation of incretins such as glucagon-like peptide-1 (GLP-1).

Glucose is primarily catabolized to generate ATP through glycolysis and oxidative phosphorylation (Figure 1). After entering cells via glucose transporters, glucose is

phosphorylated to glucose 6-phosphate (G6P), converted to fructose 6-phosphate (F6P), and then to fructose 1,6-bisphosphate (F1,6P₂) by phosphofructokinase-1 (PFK-1). This is cleaved into trioses that yield 2 ATP and 2 lactate (or pyruvate) per glucose. Subsequent oxidation of pyruvate through the TCA cycle and oxidative phosphorylation produces an additional 20–30 ATP per glucose molecule.

Glucose metabolism is tightly regulated via actions of insulin, which promotes both glucose catabolism and glucose storage as glycogen and lipids. Glycolysis is further regulated by energy balance, with ATP inhibiting and AMP and ADP promoting the key regulated step of phosphofructokinase-1. Glycolysis is feedback inhibited by key downstream carbonaceous products including lactate and citrate (Figure 1). The balance between positive and negative regulation of glucose catabolism helps maintain energy and metabolite homeostasis. Yet glucose can still have pernicious metabolic effects beyond its calories. Specifically, insulin drives fat and glycogen storage, providing an argument for the glucose-insulin model of obesity. So how is fructose different?

Fructose Metabolism.

In contrast with glucose, fructose metabolism is not tightly regulated by insulin or feedback signals. While in principle, fructose can be metabolized by hexokinase to generate fructose 6-phosphate (F6P) with further catabolism by the same glycolytic pathway as glucose, in practice fructose mainly follows an alternative pathway (Figure 2). This alternative pathway begins with fructokinase (also known as ketohexokinase, KHK), which has higher affinity for fructose than do standard hexokinases. Crucially, KHK phosphorylates fructose on carbon one, making fructose 1-phosphate (F1P), a distinctive metabolite with signaling effects. KHK has two major isoforms. KHK-C has high affinity ($K_m = 0.5$ mM) for fructose and is expressed in small intestine, liver, and kidney, as well as brain microglia and pancreatic islet cells^{9,10}. KHK-A is lower affinity ($K_m = 10$ mM) and is ubiquitously expressed but at low levels. Thus, most fructose is metabolized by KHK-C⁹.

The resulting F1P, in addition to serving as a signal, can be broken down by the enzyme F1P aldolase (also known as aldolase B) without further phosphorylation into three carbon units, bypassing the heavily regulated glycolytic enzyme PFK-1 (Figure 2). One of these three carbon units is a standard lower glycolytic intermediate (dihydroxyacetone phosphate) and the other is free phosphorylated glyceraldehyde, which has multiple potential fates: phosphorylation by triose kinase to enter lower glycolysis, reduction to glycerol, or oxidation to glycerate (see Figure 3). Consequences of the distinctive metabolic pathway for fructose include production of the pro-lipogenic signaling metabolite F1P; flow of fructose carbon into F1P, and inhibition of PFK-1 in the classic glycolytic pathway, and generation of lipogenic substrates without the positive and negative regulatory signals that normally tune glycolytic flux to energy status; and thus promotion of *de novo* lipogenesis (DNL).¹¹

Fructose-associated signaling and non-caloric effects

1] Rapid ATP Consumption, Nucleotide Degradation and Uric Acid Generation. The phosphorylation of fructose to F1P by KHK-C is rapid and results in a transient drop in ATP concentration in the liver (Figure 4)^{12,13}. While seemingly paradoxical – an energy

substrate depleting ATP – this reflects initial ATP-consuming steps outpacing downstream ATP-generation. This occurs because of the rapidity of the reaction and because there is no regulatory feedback to slow the reaction to preserve ATP levels.

Fructose-induced ATP depletion has been demonstrated in the livers of subjects taking fructose orally (75 g) as measured by magnetic resonance spectroscopy¹⁴, as well as intravenously using lower doses^{15,16}. ATP typically recovers within one hour. This effect is most severe in pericentral hepatocytes and is not observed in mammals similarly administered glucose.¹⁷

The fall in ATP levels is accompanied by a rise in adenosine monophosphate (AMP) and fall in guanosine triphosphate (GTP) and inorganic phosphate (Pi) levels (see Figure 4). The low GTP and Pi levels activate liver and intestinal AMP deaminase-2 (AMPD2) to convert the AMP to inosine monophosphate (IMP)^{13,18}. The IMP is further degraded, generating uric acid. Both intracellular and serum uric acid levels rise rapidly—within 15 to 60 minutes—following fructose ingestion, coinciding with a drop in intracellular ATP levels¹⁹. While the acute postprandial rise in serum uric acid lasts only a few hours, both fasting and postprandial serum uric acid levels rise when fructose is administered long-term²⁰. This is mediated by stimulation of urate synthesis²¹.

2| Unregulated Fructolysis. F1P is cleaved by aldolase B to generate trioses that enter into glycolysis or gluconeogenesis but bypass PFK-1, which is the site where glycolysis is normally regulated²². This allows for the rapid and unregulated production of glycolytic end products (see Figure 3). Isotope studies document as much as 25% of fructose is converted to lactate; other major fructose fates are resynthesis into glucose/glycogen, glycerate generation, and oxidation in the liver²³.

3| F1P as a Signaling Molecule that Enhances Glucose Metabolism. F1P accumulates when KHK makes F1P faster than F1P is cleaved by aldolase B. Such accumulation is augmented by an inhibition of aldolase B by the IMP²⁴. In the setting of fructose consumption, the accumulated F1P functions as a nutrient-sensing signal. One of the signaling effects of F1P is activation of liver glucose catabolism via glucokinase, the low-affinity liver-specific hexokinase. F1P dissociates glucokinase from its regulatory protein (GCKR) in the hepatocyte nucleus. Glucokinase then translocates to the cytosol where it stimulates liver glucose uptake and production of G6P that can feed glycogen synthesis or the pentose phosphate pathway; however, glycolysis is attenuated due to PFK-1 inhibition (Figure 2).²⁵ The result is a buildup of these hexoses that activates the polyol pathway.

Thus, in addition to being made from dietary fructose, F1P may also arise from an alternative pathway of glucose catabolism, in which glucose is first converted to fructose before being broken down. Hepatic flux from glucose to F1P has been demonstrated by isotope tracing.²⁶ The first enzyme of this pathway, aldose reductase (AR), has a similar affinity for glucose as GCK and hence this pathway may be substantial in the liver (Figure 2). Thus, in addition to directly being catabolized into pyruvate/lactate, fructose can also promote liver glucose catabolism.

4] ChREBP Orchestrates Fructose-mediated Events. Fructose rapidly activates the carbohydrate-sensing transcription factor (ChREBP) and induces expression of the ChREBP- β isoform in the intestine and liver^{6,27,28}. In the liver, ChREBP may be activated by metabolites produced from fructose like glycerol 3-phosphate²⁹ or xylulose 5-phosphate³⁰. It may also be triggered by fructose's induction of uric acid³¹ and of glucose-derived metabolites whose levels rise in response to glucokinase activation by F1P²⁸. ChREBP drives many fructose-dependent pathways, including glycolysis, fructolysis, gluconeogenesis, and *de novo* lipogenesis (DNL) (see Figures 3)²⁸. DNL is stimulated by fructose to a greater extent than glucose³²⁻³⁵ and this is mediated in part by ChREBP²⁸ and the sterol regulatory element-binding protein (SREBP1c)³². Activation of both transcriptional pathways is greater with fructose than glucose³².

Cellular and Physiological role of Fructose signaling

1] Shift from Fatty Acid to Carbohydrate Oxidation. Fructose ingestion causes a remarkable shift from fatty-acid to carbohydrate oxidation as measured by an increase in the respiratory exchange ratio (RER) that is greater than that observed with glucose³⁶⁻³⁸. This occurs despite glucose more strongly inducing circulating insulin, a major effector of increased RER. The decrease in fatty-acid oxidation following fructose ingestion is distinct from glucose and is greatest in the postprandial period^{36,37}, and may be mediated, in part, by fructose's ability to increase hepatocellular malonyl CoA, which drives fatty acid synthesis and inhibits fat oxidation (see Figure 3)^{25,38}. However, inhibition of hepatic fatty acid oxidation is likely insufficient to account for this acute, systemic shift to carbohydrate oxidation and reduced fatty acid oxidation in skeletal muscle is also likely required³⁶.

2] De Novo Lipogenesis (DNL). Several mechanisms have been identified for how fructose metabolism stimulates DNL in the liver. First, fructose metabolism is associated with citrate accumulation, perhaps by inhibition of mitochondrial aconitase. Citrate is a substrate for ATP citrate lyase (ACLY) and subsequent DNL (Figure 3)³⁹. Excess fructose intake can also lead to passage of fructose to the colon, where it can be metabolized to acetate, that subsequently can reach the portal vein and liver, to be converted to acetyl CoA and fat⁴⁰. Fructose metabolism in the intestine also causes local ATP depletion, F1P accumulation, and ChREBP activation^{27,41}, which may disrupt tight junctions and cause leaky gut⁴². This leakiness can produce endotoxemia that stimulates cytokine release by hepatic macrophages and Kupffer cells that can contribute to upregulation of enzymes involved in DNL⁴³. Suppression of endotoxemia with antibiotics in fructose-fed mice reduces hepatic fat accumulation, as can blocking KHK metabolism specifically in the liver⁴⁴. This suggests that the fatty liver induced by fructose is dependent on multiple mechanisms including substrate availability, enzymatic activity, and inflammation.

3] Increased Circulating Triglycerides. Fructose consumption elevates circulating triglycerides, which typically peak 4 to 6 hours after ingestion. The magnitude of triglyceride increase depends on the dose and duration of fructose exposure and is associated with elevated postprandial serum apolipoprotein B^{34,45} and in the long-term with increased visceral obesity³⁴. These effects are not observed to the same extent with equivalent glucose consumption^{34,45}.

Whether or not fructose carbons are incorporated into the circulating triglycerides is a matter of debate. Fructose increases the synthesis of triglycerides as noted by studies using labeled fatty acids (palmitate) or fructose³⁷. One isotope tracer study found that nearly 40 percent of the glycerol fraction of triacylglycerol contained fructose carbons (probably from the glycerol-3-phosphate generated during fructose catabolism, see Figure 2), while only a small amount of fructose carbon (<1%) were present in the fatty acids of the VLDL-triacylglycerol³⁷. However, fructose carbons are incorporated into lysophosphatidylcholines, and these lipid species have not been the focus of prior isotopic tracing studies⁴⁶. Thus, fructose contributes to DNL both through substrate provision and lipogenic signaling.

Other mechanisms for the increase in circulating triglycerides have been identified. First, fructose-containing SSBs increases intestinal absorption of fats in mice in part by lengthening villi⁴⁷. Metabolism of fructose also results in impaired clearance of VLDL-triglycerides⁴⁸, possibly by altering lipolysis in adipose, or by decreased removal of VLDL-triglycerides by the liver³⁷ that is linked with a fructose-dependent rise of apolipoprotein CIII⁴⁹.

4| Hepatic and Systemic Insulin Resistance. While dietary fructose does not elicit an insulin response due to lack of the fructose transporter, GLUT5, on the islet cells, over time there is development of both hepatic and peripheral insulin resistance and associated hyperinsulinemia^{32,34}. In contrast glucose-enriched diets induce hepatocellular insulin resistance to a lesser degree³². Hepatic insulin resistance is linked with fructose-induced steatosis and accumulation of specific lipid species like ceramides⁵⁰, a reduction in insulin signaling due to a reduction in insulin receptor and insulin-receptor substrate-2 (IRS2) expression, increased protein-tyrosine phosphatase 1B (PTP1b),³² and inhibition of the PI3K/Akt signaling pathway³². In addition, fructose-induced glucose production is enhanced by ChREBP-mediated upregulation of G6P phosphatase (G6Pase), the final step in gluconeogenesis²⁸. Hepatic insulin resistance in humans has been reported two to three weeks after ingestion of fructose-based beverages, but not glucose-based beverages^{51,52}.

Systemic insulin resistance has been documented with fructose feeding in rats, with impaired insulin signaling in skeletal muscle and adipocytes⁵³. In skeletal muscle there is impaired translocation of the glucose transporter, GLUT4, to the plasma membrane, and some reports of impaired insulin mediated Akt phosphorylation, possibly related to skeletal muscle accumulation of advanced glycation end products (AGEs)^{54,55}. Similarly impaired Akt phosphorylation also develops in adipose tissue of fructose-fed rats⁵³. There is evidence that the peripheral insulin resistance may develop secondary to events downstream of KHK-mediated fructose metabolism in the liver.^{56,57} Similarly, the administration of fructose to humans alters fat oxidation and mitochondrial function in skeletal muscle⁵⁸, and, similar to rats, hepatic insulin resistance occurs first followed by systemic insulin resistance^{51,59}.

Of interest, cerebral insulin resistance develops in response to dietary fructose manifesting as reduction in the signaling of the insulin receptor and IRS1⁶⁰. The cerebral effects appear to be mediated by local fructose metabolism by microglia expressing KHK¹⁰.

5] Biologic Effects from Nucleotide Degradation. Activation of the nucleotide-degradation pathway by fructose may lead to biological consequences beyond altered nucleotide levels. These may include the stimulation of intracellular and mitochondrial oxidative stress, which can occur via uric acid-mediated translocation of NADPH oxidase to the mitochondria³⁹. Uric acid and associated oxidative stress may also impact the activity of other key metabolic enzymes and regulatory factors, including inhibiting aconitase in the TCA cycle^{50,61} and enoyl CoA hydratase, an enzyme in β -oxidation³¹. A rise in intracellular uric acid can also activate ChREBP³¹ and SREBP-1c⁶², as well as stimulate expression and activation of KHK³¹ and AR,⁶³ which can lead to further fructose generation (Figure 4). Uric acid also inhibits AMP-activated protein kinase (AMPK)⁶⁴, and stimulates proinflammatory and vasoconstrictive factors⁶⁵.

6] Effects of Fructose on Mitochondrial Function. Studies in fructose-fed animals document that fructose-induced ATP consumption is commonly associated with a compensatory stimulation of the TCA cycle and enhanced mitochondrial respiration and coupling^{26,50,66}. Mitochondrial oxidative stress due to translocation of NADPH oxidase to the mitochondria also occurs^{10,39} associated with downregulation of nuclear factor erythroid 2-related factor-2 (NRF2) and antioxidant pathways⁵⁰. Possibly as a consequence, long-term consumption in rats of high-fructose corn syrup has been reported to adversely affect liver mitochondrial function, with decreased mitochondrial numbers, disrupted cristae, reduced ATP production and decreased energy efficiency^{61,67,68}. While studies in humans remain limited, there is some evidence that fructose impairs mitochondrial function in muscle⁵⁸. Accordingly, reducing fructose intake could potentially enhance mitochondrial health⁶⁹.

7] Vasopressin Regulation. Much like glucose stimulates a hormonal response in the form of insulin, fructose stimulates production and release of the hormone vasopressin, also known as antidiuretic hormone⁷⁰. Vasopressin is synthesized in the hypothalamus and released by the pituitary to increase water retention and blood pressure. The rise in plasma vasopressin levels in humans in response to fructose does not occur with equimolar glucose⁷⁰. Studies in mice show that the release is dependent on fructose metabolism by KHK⁷¹. SSBs also increase vasopressin (as reflected by an increase in plasma copeptin, a stable cleavage product of the vasopressin pro-protein)⁷², and circulating copeptin levels both predict and are elevated in subjects with metabolic syndrome and diabetes⁷³.

The arginine vasopressin receptor 1a (V1a) receptor is responsible for the increased systemic blood pressure triggered by vasopressin. In contrast, the V1b receptor mediates vasopressin-induced production of adrenocorticotropic hormone (ACTH). ACTH stimulates the release of cortisol and glucagon, powerful glucose-elevating hormones. Mice lacking vasopressin V1b receptor (but not V1a receptor) have lower cortisol, glucagon, and liver KHK and are protected from fructose-induced metabolic syndrome⁷¹. This metabolic improvement may not be specific to fructose, as mice lacking V1b show reduced ACTH and corticosterone responses in response to other stressors such as forced swimming. The role of vasopressin suggests that better hydration, which lowers vasopressin, could potentially mitigate metabolic syndrome, with proof-of-concept shown in mice⁷¹. Thus, vasopressin is a fructose-induced hormone with strong connections to blood pressure and metabolic

syndrome. Both the ancient reasons for evolution of the fructose-vasopressin circuit and the clinical potential of its targeting to treat modern metabolic problems merit further investigation.

DIETARY FRUCTOSE ABSORPTION AND METABOLISM

Table sugar (sucrose) is a disaccharide containing equimolar fructose and glucose, whereas high fructose corn syrup (HFCS) is most commonly a 55:45% mixture of fructose and glucose. The major source of dietary fructose comes from SSBs and ultra-processed foods with abundant sucrose or HFCS, but fructose is also high in fruits, fruit juices, and honey. Small amounts of fructose are also present in certain vegetables (such as sweet potatoes and carrots).

Whereas HFCS sugars are ingested as monosaccharides, the sucrose disaccharide is cleaved into glucose and fructose in the small intestine by sucrase-isomaltase. Despite this extra enzymatic step, there appears to be minimal difference in the physiological effects of sucrose and HFCS⁷⁴. Glucose is absorbed by enterocytes in the small intestine by the sodium dependent transporter SGLT1, while fructose is taken up by GLUT5, and both exit into the portal venous system through the GLUT2 transporter.

Once in the enterocyte, fructose is metabolized by KHK-C, generating primarily glucose and organic acids (Figure 5).⁷⁵ With low intake, most fructose is metabolized in the intestine, but higher intakes can saturate intestinal metabolism and lead to greater fructose delivery to the liver. Thus, intestinal metabolism functions to shield the liver from small quantities of ingested fructose.⁷⁵ The fructose clearance rate of the human intestine is uncertain. Fructose metabolism in the gut also enlarges intestinal villi, increases absorption of high-fat foods, and may induce leaky gut by disrupting tight junctions.

Fructose absorption is affected by many factors. For example, it is enhanced by the presence of glucose via mechanisms still not fully understood⁷⁶. Expression of GLUT5, the essential luminal fructose transporter, is low in infancy, but expression increases in response to dietary fructose and sugar⁷⁷. This increase is dependent on ChREBP²⁷. Higher GLUT5 expression is associated with a higher risk for obesity in children⁷⁸.

Fructose-based drinks are also more effective than fructose-based foods to induce KHK-dependent metabolism and downstream events in the liver⁷⁹. Not only is this due to the amount ingested, but the speed of ingestion⁷², likely because rapid intake leads to saturation of intestinal fructose clearance and higher fructose concentrations in the liver. The presence of fiber can also slow absorption, and, along with ascorbate, potassium, and flavanols, slow or inhibit fructose metabolism. This may help explain why ingestion of fruit, the main source of natural fructose, does not typically result in adverse metabolic effects^{8,80}. Ingestion of inulin (a fructan polymer consisting of a linear chain of fructose molecules) can stimulate the growth of bacteria that will degrade fructose in the gut and protect mice from fructose-induced fatty liver⁸¹. However, other fructans (such as levans that form net-like structures of fructose) can be degraded by bacteria such as *Streptococcus* that releases fructose, and may induce metabolic syndrome, at least in horses⁴².

Of the dietary fructose that reaches the liver, the majority is metabolized on first pass. Many of the detrimental metabolic effects of fructose are mediated by its metabolism in the liver, and liver-specific KHK knockout mice are protected from fructose-induced fatty liver, insulin resistance, and obesity⁵⁷. Only 10 to 20 percent of the ingested fructose enters the systemic circulation^{6,75,82}, where it is metabolized by other tissues (with contributions from the kidney, muscle, heart, adipose tissue, lung, brain, and circulating monocytes^{9,10,83-85}) or excreted in the urine. Overall, the first-pass metabolism of fructose aligns with its role as an ancient signal to store carbohydrate as fat, with intestinal clearance ensuring that the liver 'fat switch' is turned on only when fructose is sufficiently abundant.

ENDOGENOUS FRUCTOSE PRODUCTION AND METABOLISM

Fructose can be converted to glucose via gluconeogenesis where fructolytic products are diverted to F1,6P₂, F6P, G6P and then to glucose (see Figure 3). Conversely, glucose can be transformed to fructose via the *polyol pathway*: glucose is first converted to sorbitol by aldose reductase (AR) followed by conversion to fructose by sorbitol dehydrogenase (SDH) (see Figure 4).

Constitutive production of fructose has been documented in human reproductive organs including in the placenta in early pregnancy where fructose is present in amniotic fluid⁸⁶ and in the prostate gland and seminal vesicles where fructose concentrates in semen⁸⁷. Sperm use hexokinase and not KHK to metabolize fructose⁸⁷, and both mice and humans lacking KHK are fertile.

AR, the committal enzyme, is not normally expressed in non-reproductive tissues, except for the renal medulla where it generates sorbitol as an intracellular osmolyte that protects kidney epithelial cells from osmotic stress. Importantly however, AR can be induced in other tissues by stimuli including hyperglycemia, hyperosmolarity, hypoxia, ischemia, heat, trauma, hyperuricemia, and glucokinase activation (Figure 5 and Table 2)⁸⁸⁻⁹⁷.

Historically, the role of endogenous fructose was considered minimal. However, a study in mice found that chronic glucose ingestion led to obesity and metabolic syndrome accompanied by induction of AR with accumulation of fructose and sorbitol in the liver⁹⁸. Importantly, mice lacking either AR or KHK were protected from developing fatty liver or insulin resistance⁹⁸.

Endogenous fructose production is increased with high salt⁸⁸ or alcohol intake⁹⁹ and this endogenously produced fructose may contribute to metabolic syndrome and fatty liver in these contexts. Experimental studies have now identified increased endogenous fructose production in many contexts, including diabetes, ischemic or heat-stressed kidneys, ischemic heart, and the brains of animals with diabetes, head trauma, dehydration, or intake of high salt or high sugar diets^{84,92,100-102}. Systemic hypoxia can also increase serum fructose levels in the naked mole rat, and this facilitates survival in hypoxic burrows, nominally by promoting glycolysis¹⁰³.

Evidence for endogenous fructose production in humans is also emerging. Francey et al documented a tripling of the endogenous fructose production rate following administration

of an oral glucose-fructose beverage in healthy people⁸². ‘Clamping’ blood glucose at hyperglycemic levels in healthy individuals induced fructose production in human brain¹⁰⁴. Interestingly, fructose and sorbitol levels are elevated in brain tissue or cerebral spinal fluid in multiple sclerosis, cranial hypertension, Alzheimer’s disease, bipolar disease, pregnancy, and individuals with brain tumors¹⁰⁵⁻¹¹¹. Various cancers have also been shown to have AR expression associated with fructose production¹¹². Diabetes, obstructive sleep apnea, and chronic kidney disease are all associated with increased fasting circulating fructose levels in human populations suggestive of increased endogenous fructose production in these conditions.¹¹³⁻¹¹⁶

These studies suggest that the potential exposure to fructose is high in western societies, not only from added sugars in the diet, but due to the high dietary content of carbohydrate that can provide substrate for fructose production, a process amplified by intake of salty foods and alcohol.

BIOLOGICAL CONSEQUENCES OF FRUCTOSE METABOLISM

An important question is how fructose metabolism translates into its biological effects. Indeed, it is the unique aspects related to FIP signaling, ATP depletion, uric acid generation, and stimulation of glycolytic and lipogenic pathways that drive the biological effects of fructose metabolism.

Taste, Hunger and Food Intake

Sugar (sucrose) and HFCS are sweet and palatable, and the sweet signals activate orexigenic pathways, including an amygdala to hypothalamic circuit that stimulates consumption of sweet foods¹¹⁷. However, the increase in food intake cannot fully explain the effects of fructose on systemic metabolism. For example, mice lacking sweet taste receptors still develop a dopamine response to sucrose whereas this does not occur with artificial sugars¹¹⁸. Indeed, mice lacking taste still seek sugar (or fructose), and, although they drink less than normal mice, they still develop metabolic syndrome¹¹⁹.

KHK knockout mice no longer prefer fructose, although they continue to prefer glucose. Mice lacking KHK only in the intestine show reduced preference for fructose, but demonstrate enhanced delivery of fructose to the liver that leads to increased propensity for metabolic syndrome^{57,75}. It is possible that mice lacking intestinal KHK may have some impaired absorption of fructose that could cause subclinical gastrointestinal side-effects that lead to less intake⁵⁷. Mice with liver-specific KHK knockout continue to show marked preference for and excessive intake of fructose, but are protected from weight gain or metabolic syndrome⁵⁷. These studies show that while the taste and palatability of fructose encourages intake, it is the metabolism of fructose in the liver that drives metabolic disease.

Central orexigenic and other pathways are also likely important for how fructose induces hunger and food intake¹²⁰. For example, glucose injected into the cerebral ventricles of mice causes a rise in brain ATP levels, induces satiety and curtails food intake¹²¹. In contrast, injection of fructose lowers ATP, stimulates hunger, and increases food intake¹²¹. Fructose ingestion also causes in humans a reduction in cerebral cortical blood flow to

brain areas involved in self-control while stimulating occipital visual centers to identify appealing foods^{122,123}, consistent with the stimulation of foraging-like activity^{124,125}. In contrast, glucose acutely increases cortical blood flow and satiety. The liver ATP depletion that occurs with fructose also stimulates hunger¹²⁶.

Nevertheless, mice provided fructose in drinking water initially compensate for the fructose caloric intake by lowering their intake of chow to maintain overall caloric balance¹²⁷. This can last for several weeks, but then food intake gradually increases with an increase in weight. Experimentally this is associated with the development of central leptin resistance.^{128,129}

While experimental studies favor fructose metabolism in driving hunger and weight gain, fructose (and sucrose) stimulates hepatic fibroblast growth factor 21 (FGF21) that acts to reduce sugar intake and sweet preference in human. Ingestion of glucose is less potent than fructose in stimulating FGF21¹³⁰. Studies in laboratory mice suggest the stimulation of FGF21 is dependent on fructose metabolism by hepatic KHK⁵⁷. These studies suggest fructose metabolism may also have a negative feedback system to limit excessive sugar intake.

Metabolic Syndrome and Obesity

The administration of fructose to mice and rats is commonly used to induce obesity, metabolic syndrome, diabetes, and fatty liver. However, large doses are often required when incorporated into food (50 to 60%) and the most effective method is to provide fructose in the drinking water as HFCS at concentrations similar to SSBs¹³¹. Studies in mice have found that the metabolic syndrome can be prevented if KHK-C is knocked out or inhibited^{32,127,132}. While this could be attributed to that fact that KHK KO mice drink less fructose than wild type controls, the protection persists when fructose consumption is matched¹²⁷.

In humans the metabolic effects of fructose are also best observed with administration of fructose in drinks rather than in food, and especially in individuals who are older, overweight or have borderline insulin resistance¹³³. One study in healthy, physically active and lean young adults found minimal effects when given packets of high dose (150 g/d) crystalline fructose daily for 8 weeks¹³⁴, while another study in overweight men found that high doses of fructose in the drinking water (200g/d) resulted in increases in blood pressure, fasting insulin and serum triglycerides within 2 weeks¹³⁵.

In a prospective clinical trial, fructose-sweetened or glucose-sweetened beverages were administered as 25 percent of the energy requirement to overweight or obese individuals for 10 weeks. Fructose supplementation promoted features of metabolic syndrome including decreased fat oxidation and insulin sensitivity and increased visceral obesity, DNL, postprandial plasma triglycerides and uric acid, and fasting apolipoprotein B, LDL cholesterol, plasminogen activator inhibitor-1 (PAI-1), and monocyte chemoattractant protein-2 (MCP-1) compared to the group receiving glucose^{20,36,136}.

While fructose induces worse metabolic effects than glucose in humans, most SSBs include both fructose and glucose. Moreover, and as mentioned earlier, experimental studies suggest

that over time glucose-sweetened beverages may increase endogenous fructose production.⁵⁷ This can potentially confound studies that attempt to distinguish effects of these two sugars.

Another confounding issue is the common cooccurrence of excess calorie intake and fructose consumption. If caloric intake is controlled, fructose consumption has a minimal effect on body weight in both experimental^{77,137} and human¹³⁸ studies. Any observed modest weight increases are likely attributable to higher expression of transporters (GLUT5) for fructose^{78,139} or greater intestinal villous area that allows more efficient absorption of fat⁴⁷.

As evidenced by pair-feeding studies, however, sugar, HFCS, glucose, or fructose can produce metabolic disease even if weight does not increase^{77,98,140,141}. For example, one study found that rats can develop diabetes, fatty liver and hypertension on a high sugar diet despite caloric restriction⁷⁷. Consistent with this observation, men consuming a weight-maintaining diet that included fructose-sweetened beverage for 9 days (versus a weight-maintaining diet with the fructose beverage) showed decreased fat oxidation and increased DNL, liver fat, postprandial triglyceride and hepatic insulin resistance⁵⁹. In another study, isocaloric restriction of fructose in obese adolescents improved features of metabolic syndrome¹⁴². Thus, even in the absence of excessive caloric intake, sugar can drive metabolic disease.

Metabolic Dysfunction-Associated Steatotic Liver Disease (MASLD) and Alcoholic Liver Disease

SSBs are strongly linked with the development and severity of MASLD, likely due to the effect of fructose to stimulate DNL and to inhibit fatty-acid oxidation¹⁴³. Subjects with MASLD have been found to have elevations of KHK mRNA in the liver biopsies¹⁴⁴. Administration of SSBs has also been found to increase liver fat^{74,145} while reducing fructose intake can decrease liver fat^{146,147}. Experimental studies have also found that knocking out KHK can substantially reduce fatty liver in mice given a high fat, high sugar diet as well as a diet supplemented with HFCS^{57,148}. Fatty liver from high glycemic or high salt diets are also associated with endogenous fructose production and are prevented in mice lacking KHK^{88,98}. These studies provide strong support for fructose as a key driver of MASLD.

Alcohol intake can also lead to fatty liver and cirrhosis with histological and biochemical similarities to MASLD. Indeed, the combination of fructose with alcohol can markedly accelerate development of liver disease⁹⁹. More recently it has been shown that alcohol-induced liver disease also involves endogenous fructose, likely resulting from induced AR due to alcohol's osmotic effects.^{99,149,150} AR is also induced in the liver of patients with alcoholic liver disease. Mice lacking AR or KHK are largely protected from alcohol-induced liver disease. Of interest, mice lacking KHK or given KHK inhibitors also drink less ethanol⁹⁹.

Hypertension, Kidney Disease, and Other Conditions

A variety of other conditions have been linked with fructose metabolism. Primary hypertension, for example, is strongly linked with intake of SSBs. There is also an

interesting interaction, as fructose enhances sodium absorption in the gut¹⁵¹ and sodium reabsorption in the kidney tubules^{151,152}, and high salt intake raises osmolality and induces endogenous fructose production⁸⁸. Perhaps most strikingly, mice lacking KHK are protected from salt-induced blood pressure elevation and cardiac hypertrophy^{88,153}.

Systemic inflammation can potentially also be induced by fructose, by effects on circulating monocytes^{83,85} and the release of chemokines into the circulation¹³⁶. Local inflammation can also be induced, such as the release of inflammatory mediators by kidney epithelial cells¹⁵⁴, or by the activation of local macrophage-like cells including Kupffer cells¹⁵⁵ and microglia¹⁰.

Acute and chronic kidney disease may also be dependent on endogenous fructose production, including in experimental models of acute ischemic tubular injury, contrast nephropathy, heat stress induced chronic kidney disease, and diabetic nephropathy^{92,100,102}. In mouse models of these conditions, endogenous fructose production was shown to occur in the kidney, and mice lacking the ability to metabolize the fructose (i.e., KHK KO mice) were protected from kidney damage.

Ingestion of SSBs also increases the risk of gout. Humans and great apes are much more susceptible to gout from fructose as they lack the hepatic enzyme, uricase, that helps regulate uric acid by degrading it to allantoin. Indeed, mice or rats that have uricase knocked out or inhibited not only have higher uric acid levels but show enhanced risk for hypertension, kidney disease and metabolic syndrome and also a relatively greater metabolic response to fructose (reviewed in¹⁵⁶).

One of the more interesting findings involved the process of aging. Mice and rats are known to develop aging-associated kidney disease, similar to humans. However, a study of aging KHK knockout mice on a high carbohydrate but sugar-free chow found that they were protected from age-associated kidney disease and hypertension¹⁵⁷. Since the dietary carbohydrates were corn, soybean and wheat, all with minimal fructose content, it is likely that the aging-associated changes in the wild-type mice resulted from endogenous fructose.

Cancer

Fructose metabolism has emerged as tumor promoter, supporting both tumor-intrinsic and systemic processes that promote disease progression. In multiple cancers, including breast, gastric, lung, hepatic, pancreatic, and prostate, fructose metabolism has been implicated in cancer growth, proliferation, and metastasis.¹⁵⁸⁻¹⁶² In several instances, fructose is produced endogenously by tumor cells via the polyol pathway, which facilitates cancer cell metabolism, migration, and resistance to apoptosis¹¹². In cases where tumors have access to fructose-rich environments, such as the intestine and liver, fructose can be directly obtained and utilized from dietary sources^{163,164}. That said, hepatocellular carcinoma loses fructolytic capacity compared to healthy liver and due to the expression of KHK-A instead of KHK-C, may contribute to expression of the oncogenic antioxidant transcription factor Nrf2^{165,166}. Dietary fructose can also promote tumor growth in organs without direct access to dietary nutrients via fructose-derived metabolites, like lactate and lipids, which are produced by the liver following first-pass metabolism⁴⁶. Within cancer cells, fructose

fuels glycolysis and DNL to support macromolecule synthesis and proliferation^{46,163,167}. Moreover, the major product of fructolysis, FIP, functions as a survival signal by increasing HIF1 α transactivation, thereby promoting adaptation to hypoxic conditions¹⁶³. Furthermore, fructose metabolism may shape the tumor microenvironment by influencing the activity of immunosuppressive cell populations^{168,169}. In sum, through multiple mechanisms that merit further investigation, fructose likely promotes cancer.

Brain Disorders

The human brain makes endogenous fructose (such as from hyperglycemia)¹⁰⁴ and also responds to dietary fructose. Ingestion of fructose stimulates foraging-like behavior with activation of occipital centers involved in food cues while suppressing blood flow to areas associated with self-control (cortex) and recent memory (hippocampus), while glucose has opposing effects^{122,123,170}. Chronic inhibition of cortical and hippocampal function from excessive ingestion or production of fructose has been postulated to increase the risk of behavioral disorders and dementia^{124,125}. High fructose and sorbitol levels in the brain occur in both bipolar disease^{106,171} and dementia¹¹¹. Microglia contain KHK-C and are activated by fructose¹⁰, and when this occurs in pregnant mice, the progeny display anxiety disorders¹⁷².

Fructose metabolism in the brain may have a role in Alzheimer's disease¹²⁵. Rats ingesting fructose chronically develop cognitive dysfunction, with cerebral insulin resistance, mitochondrial dysfunction, neuroinflammation, and eventually amyloid plaques and tau protein aggregates^{60,173-175}. Diabetic mice also have increased endogenous fructose production with cognitive dysfunction that is improved by targeted knockout of KHK-C in the hippocampus¹⁰. Intake of sugar and high glycemic foods are also risk factors for Alzheimer's disease, and this disease is associated with 5 to 6-fold higher levels of fructose and sorbitol in the brain in autopsy studies.¹¹¹

Genetic Disorders

Two rare genetic disorders of fructose metabolism are *essential fructosuria* (KHK deficiency) and hereditary *fructose intolerance* (HFI, aldolase B deficiency). The former is asymptomatic, while the latter results in a pathological reaction to fructose, characterized by hypoglycemia and lactic acidosis acutely and MASLD chronically (Box 1)¹⁷⁶⁻¹⁷⁸. Both experimental and pilot clinical studies suggest that inhibition of KHK could be a promising therapy for subjects with HFI^{177,179}. There are also ongoing experimental studies to genetically edit some of the common mutations that cause HFI in the hope of a permanent cure (M Lanaspá and D Tolan, unpublished data).

CONCLUSIONS AND FUTURE DIRECTIONS

Open questions

The role of endogenously produced fructose in metabolic disease remains controversial. While the risk of SSBs on metabolic health is robust, there is less evidence that fructose from other sources carries risk. Interpretation of epidemiological studies is confounded by

fruits, which are rich in fructose but carry countering substances. There are also only limited studies in humans to determine the contribution of endogenous fructose.

The temporal relationship between sugar intake and metabolic disease trends remains debated. If fructose and sugar are important drivers of metabolic syndrome, then why has decreased intake of added sugars during the last two decades failed, prior to the common use of GLP-1 agonists, to slow the obesity and diabetes epidemic? Historically, there has been a 10–20 year lag between sugar exposure and metabolic outcomes¹⁸⁰, and changes in incidence often precede changes in prevalence. Indeed, while the prevalence of diabetes has not decreased, the incidence of diabetes began to fall soon after sugar reduction¹⁸¹, while obesity prevalence plateaued around 2020 and began decreasing by 2023¹⁸². Importantly, while recent changes in obesity prevalence are impacted by widespread GLP-1 therapy, the decrease in diabetes incidence predates this.

The role of uric acid as a mediator of fructose's metabolic effects is contentious. Experimental studies link fructose metabolism to increased uric acid and metabolic dysfunction, but Mendelian randomization and large clinical trials do not support uric acid as a causal mediator of cardiovascular or renal disease. These discrepancies may stem from genetic studies measuring serum rather than tissue uric acid, and from trial designs that inadequately capture tissue-specific effects¹⁵⁶.

Finally, whether targeting fructose metabolism through KHK inhibition offers therapeutic benefit remains uncertain. Two pharmaceutical companies abandoned KHK inhibitor programs after phase 2 studies yielded modest outcomes^{183,184}. Pfizer's PF-06835919 initial studies produced only mild reductions in liver fat (~20%) without significant improvements in insulin, uric acid, inflammation, or weight. These limited benefits may reflect a short duration of treatment, suboptimal dosing, off-target effects, the need to use it in a more select population (such as those with high dietary intake of fructose), and also the possibility that KHK inhibition is more effective for disease prevention than treatment¹⁸⁵. As mentioned earlier, there is some clinical evidence that KHK inhibition may benefit subjects with HFI¹⁷⁹. Given KHK's broad metabolic role, its inhibition may still hold future promise for improving metabolic health.

Conclusion

In conclusion, fructose has multiple distinctive metabolic effects beyond standard carbohydrate metabolism. It produces signals that promote hepatic glycolysis and lipogenesis. This pro-lipogenic response presumably reflects an ancient evolutionary adaptive program intended to prepare for periods of food scarcity (Figure 6). Fructose's induction of vasopressin may similarly reflect preparation for water scarcity, with fat storage also supplying metabolic water¹⁸⁶. In modern society, however, this ancient signaling role can backfire. In the context of consistently abundant food, fructose intake is a hazard, promoting insulin resistance, hypertriglyceridemia, fatty liver, and elevated blood pressure⁸.

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Box 1**Genetic Disorders of Fructose Metabolism*****Essential Fructosuria***

- Fructokinase (KHK) Deficiency; Autosomal recessive (< 1 per 130,000).
- Normal lifespan, Asymptomatic.
- Prefer salty over sweet foods.
- May be protective against obesity or type 2 diabetes (no reports of obesity of type 2 diabetes in affected individuals)
- Excrete 10 percent of ingested fructose load in urine. Most ingested fructose is metabolized in adipose and other tissues via other hexokinases.

Hereditary Fructose Intolerance

- Aldolase B deficiency; Autosomal recessive (1:22,000).
- Manifests after weaning with exposure to fructose.
- Presents with acute hypoglycemia upon fructose intake, due to impaired hepatic gluconeogenesis.
- Biochemically, FIP accumulates pathologically, ATP is depleted.
- Causes chronic liver disease and kidney disease (Fanconi syndrome).
- Rescued by inhibition or knocking out KHK.
- Management consists of dietary fructose and sorbitol avoidance.
- Associated with fructose, sorbitol and ethanol aversion.
- Restriction of high glycemic carbohydrate to reduce endogenous fructose generation may also be helpful.

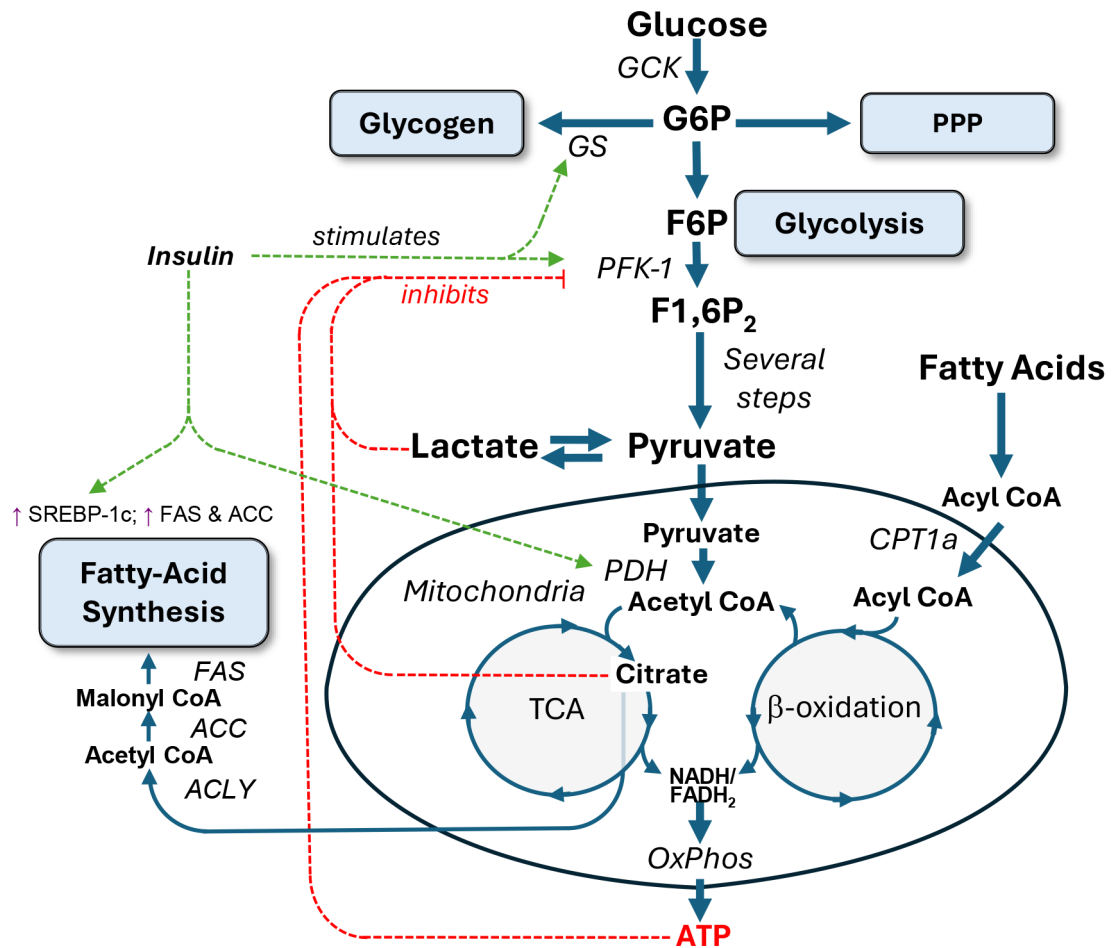


Figure 1. Hepatocyte Glucose Metabolism.

The schematic highlights insulin-stimulated (green dashed arrows) hepatic metabolic flux in the fed state that promotes energy storage. This is balanced by inhibition of glucose usage when ATP flux is high (red dashed arrows). The available pathways are denoted with light-blue boxes, the important enzymes in italics, and metabolic intermediates in bold. Abbreviations in alphabetical order are as follows: ACC, acetyl-CoA carboxylase; ACLY, ATP-citrate lyase; CoA, Coenzyme-A; CPT1a, carnitine palmitoyltransferase-1a; FAS, fatty acid synthase; F6P, fructose 6-phosphate; F1,6P₂, fructose 1,6-bisphosphate; FADH₂, flavin adenine dinucleotide (reduced); G6P, glucose 6-phosphate; GCK, glucokinase (hexokinase IV); GS, glycogen synthase; NADH, nicotinamide adenine dinucleotide (reduced); OxPhos, oxidative phosphorylation pathway; PDH, pyruvate dehydrogenase complex; PFK-1, phosphofructokinase-1; PPP, pentose phosphate pathway; SREBP-1c, sterol regulatory element binding protein-1c; TCA, tricarboxylic acid cycle.

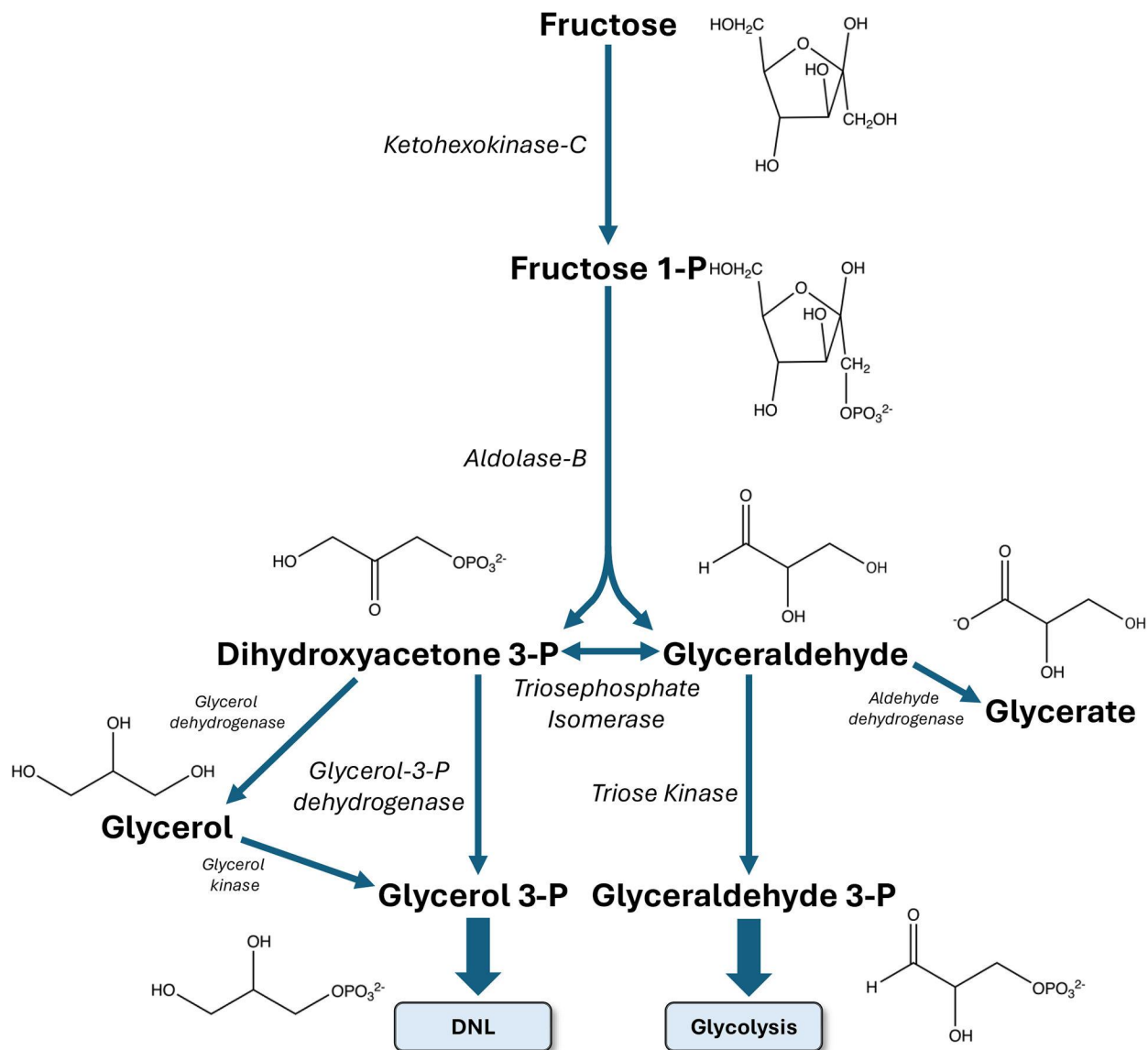


Figure 2. Fructose Metabolism.

The chemical pathway from fructose to triose phosphates is diagrammed with all known end products from fructose. The structures of metabolic intermediates are shown with names in bold. The structures of the hexoses are in the β -anameric forms utilized by the enzymes, which are noted in italics. Glycerate enters glycolysis as 2-phosphoglycerate via glycerate kinase (not shown). The major pathways, glycolysis and *de novo* lipogenesis (DNL) to which these triose phosphates enter (thick arrows) are noted by gray boxes.

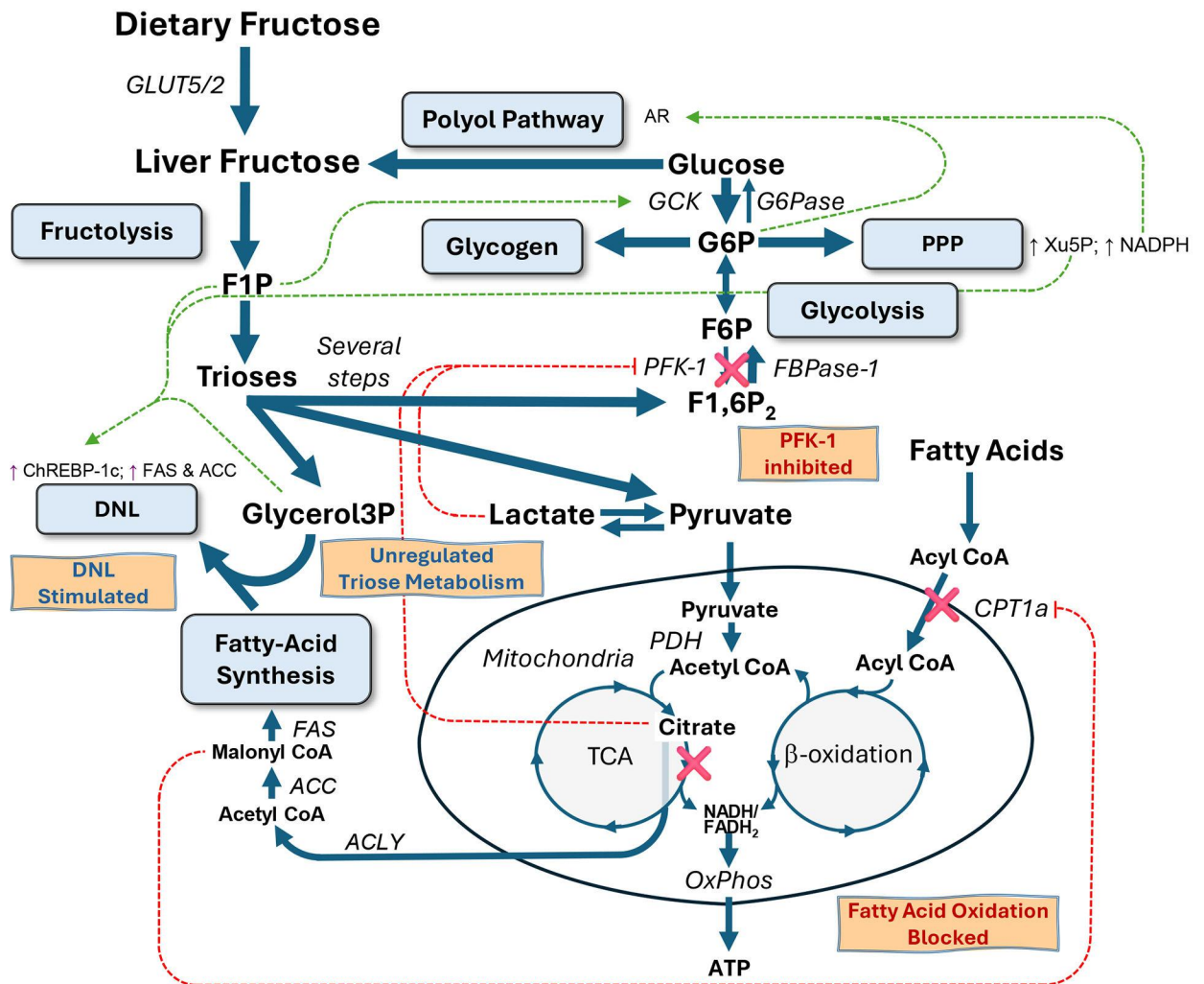


Figure 3. Hepatocyte Fructose Metabolism.

The major differences with glucose metabolism are highlighted in the colored boxes, which are consequences of regulatory effects that activate (green dashed arrows) or inhibit (red dashed arrows) key enzymes or transcription factors as indicated. This regulation leads to inhibition of these pathways as indicated with red X's. The stimulation of glucokinase (*GCK*) facilitates enhances glucose uptake and might further enhance fructose generation via the polyol pathway. The impacted pathways are denoted with gray boxes, the important enzymes in italics, and metabolic intermediates in bold. Other abbreviations in alphabetical order are as follows: *ACC*, acetyl-CoA carboxylase; *ACLY*, ATP-citrate lyase; *AR*, aldose reductase; *ChREBP*, carbohydrate-response element binding protein; *CoA*, Coenzyme-A; *CPT1a*, carnitine palmitoyltransferase-1a; *DNL*, *de novo* lipogenesis; *FAS*, fatty acid synthase; *F1P*, fructose 1-phosphate; *F6P*, fructose 6-phosphate; *F1,6P₂*, fructose 1,6-bisphosphate; *FADH₂*, flavin adenine dinucleotide (reduced); *FBPase-1*, fructose 1,6-bisphosphatase-1; *GCK*, glucokinase (hexokinase IV); *G6Pase*, glucose 6-phosphatase; *G6P*, glucose 6-phosphate; *GLUT*, solute carrier family or glucose transporter; *NADH*, nicotinamide adenine dinucleotide (reduced); *NADPH*, nicotinamide adenine dinucleotide phosphate (reduced); *OxPhos*, oxidative phosphorylation pathway; *PDH*,

pyruvate dehydrogenase complex; PFK-1, phosphofructokinase-1; PPP, pentose phosphate pathway; SREBP-1c, sterol regulatory element binding protein-1c; TCA, tricarboxylic acid cycle; Xu5P, xyulose 5-phosphate.

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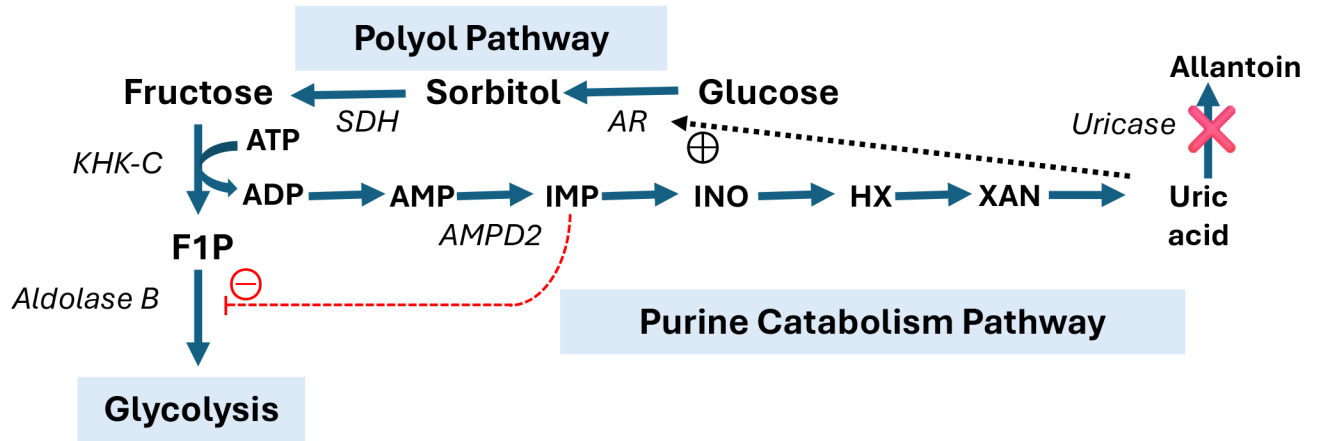


Figure 4. Fructose Induced ATP Depletion and Uric Acid Generation.

The schematic highlights the relationship among the fructolytic, polyol, and purine degradation pathways (gray boxes). The relative fluxes are indicated by thickness of arrows with the results on metabolites indicated with up or down arrows. Activation (green dotted arrows) and inhibition (red lines) of key enzymes or transcription factors are indicated. The rapid consumption of ATP by ketohexokinase-C (KHK-C) and resulting sequestration of phosphate in fructose 1-phosphate (F1P) results in transient ATP depletion and activation of AMP deaminase-2 (AMPD2) to eventually generate uric acid. Uric acid stimulates aldose reductase (AR), KHK-C, and the transcription factors sterol regulatory element binding protein-1c (SREBP-1c) and carbohydrate-response element binding protein (ChREBP). Inosine monophosphate (IMP) inhibits aldolase B perhaps enhancing the amplification of F1P sequestration. Because humans lack uricase, which converts uric acid to allantoin, the uric acid response is greater in humans than most mammals. Other abbreviations in alphabetic order: ADP, adenosine diphosphate; AMP, adenosine monophosphate; GTP, guanidylate triphosphate; P_i, inorganic phosphate; SDH, sorbitol dehydrogenase.

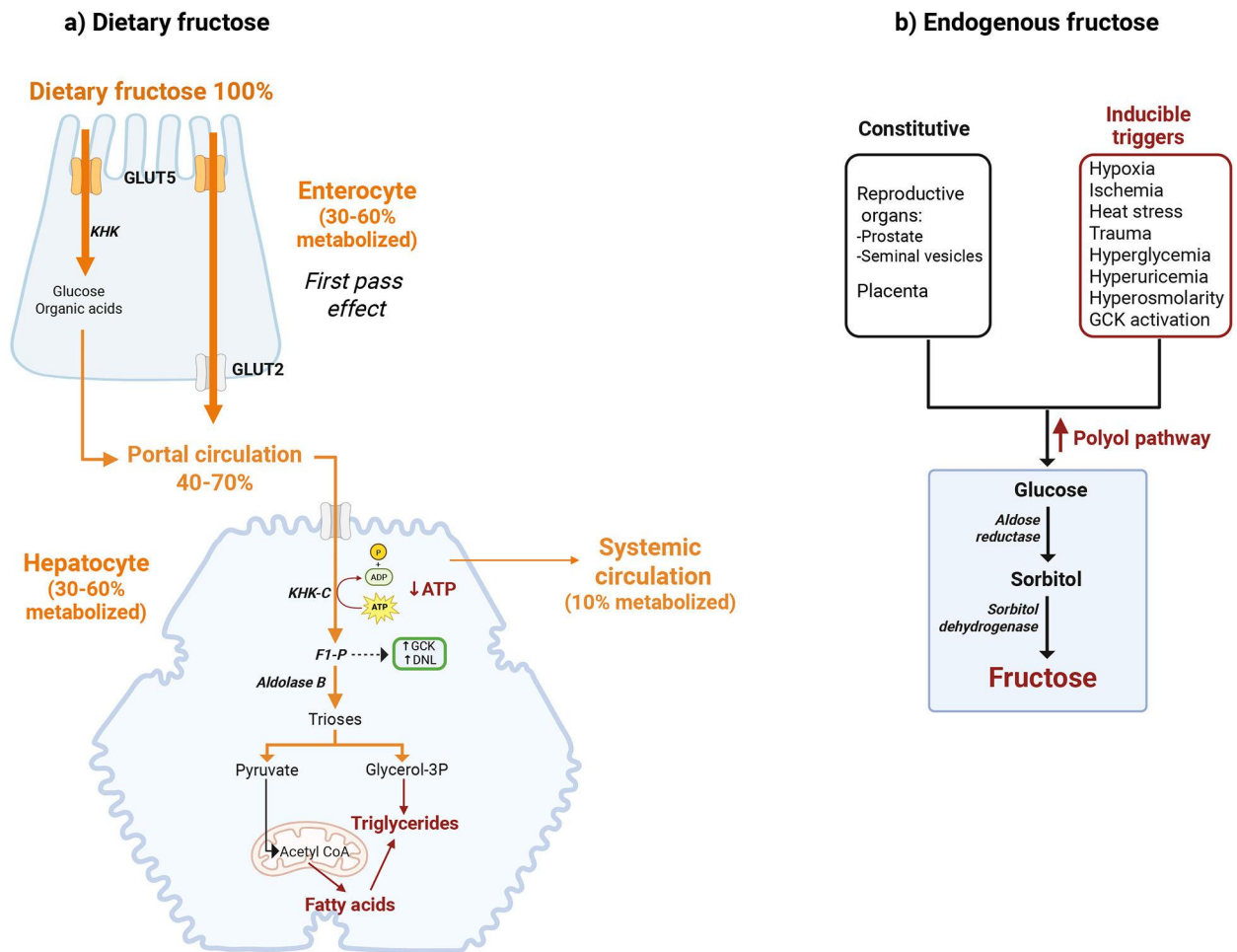


Figure 5. Sites of Metabolism for Fructose.

A) Dietary Fructose: Fructose metabolism begins with absorption by enterocytes of the small intestine, gets exported to the portal circulation, and picked up by the liver and systemic circulation. *B) Endogenous Fructose:* Fructose can also be produced endogenously from glucose through the polyol pathway (blue box). Fructose production is regulated by Aldose reductase. Those tissues with constitutive expression are listed in black in box on the left. Other tissues require induced expression by various stresses listed in brown in middle box. Created in BioRender. Sánchez-Lozada, L.G. (2025)

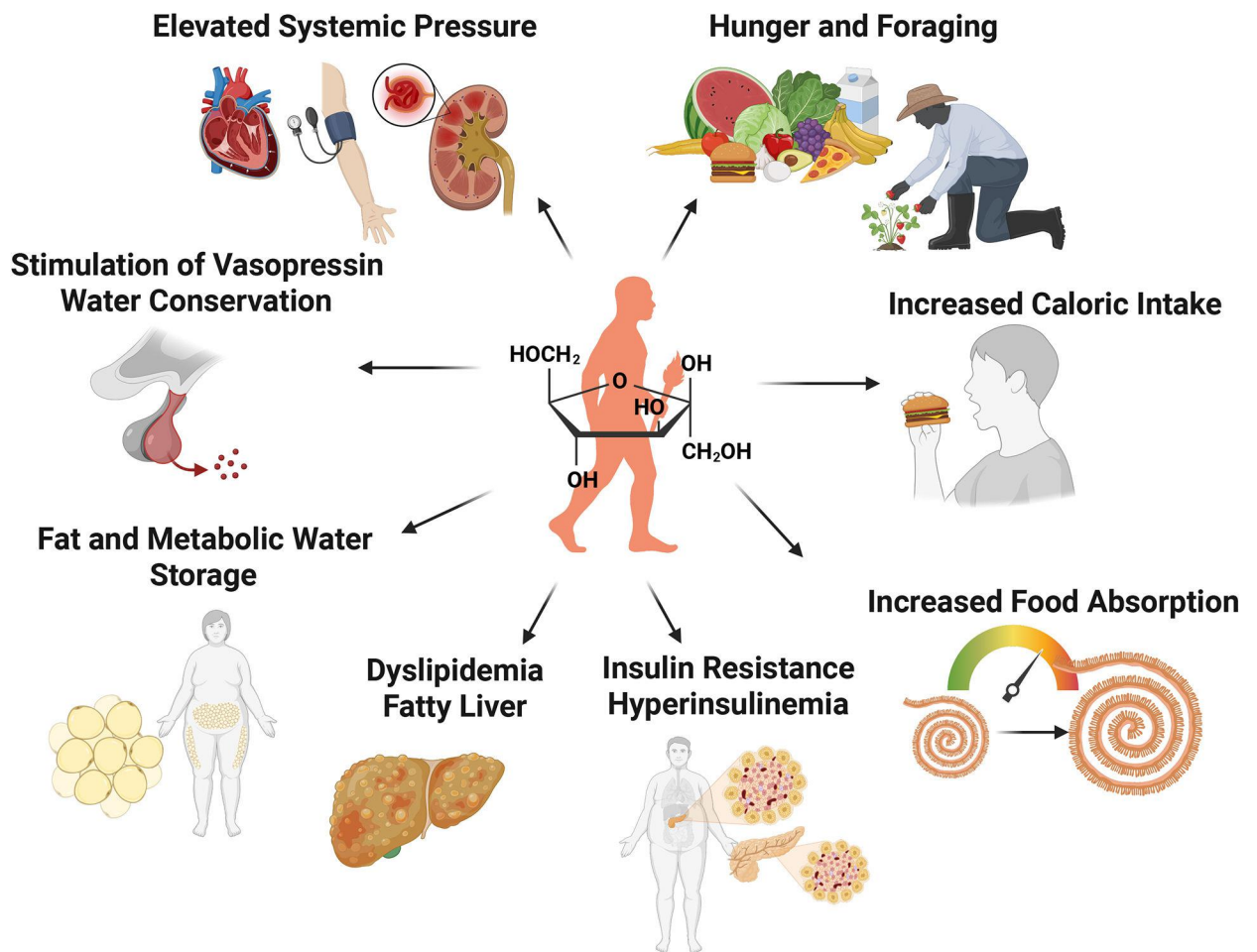


Figure 6. The Fructose Survival Hypothesis.

Fructose likely provided an evolutionary advantage by activating a “survival pathway” that increased hunger, stimulated foraging, and promoted efficient fat storage, traits that helped early hominids endure periods of famine. In modern environments with constant food availability, this once-beneficial mechanism can contribute to metabolic disease. This concept is described extensively in the literature^{8,125} and has been proposed as the “fructose survival hypothesis Created in BioRender. Lanasp, M. (2025) <https://BioRender.com/7oh5q9r>.

Table 1**Differences in Fructose and Glucose Actions**

Glucose Effects Substantially Greater than Fructose
Insulin Secretion
Stimulate satiety
Fructose Effects Substantially Greater than Glucose
Increased Carbohydrate Oxidation ³⁷
Impaired Fatty Acid Oxidation ²⁵
Increased De novo Lipogenesis ^{25,32}
Increased Circulating Triglycerides (VLDL-triglycerides) ^{25,28}
Increased Lactate production ²⁵
Acute ATP depletion ¹²¹
Rapid Uric Acid Generation ¹⁸⁷
Induce Hyperinsulinemia and Insulin Resistance ^{28,32}
Stimulate Gluconeogenesis (Increase G6P phosphatase) ²⁸
Increase Intestinal Surface (Villi) Area to Enhance Fat Absorption ⁴⁷
Induce Leptin Resistance ¹²⁸
GCK Activation ²⁸
Vasopressin Release ⁷⁰
FGF21 Release ¹³⁰
Increased Systemic Blood Pressure ¹⁸⁸
Increase Glomerular Hydrostatic Pressure ¹⁸⁹
Leaky Gut and Endotoxemia ⁴³
Reduce Satiety

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Table 2

Factors that Activate Endogenous Fructose Production

Type of Stress	Examples
Hyperosmolarity	Water Deprivation ¹⁰²
	Salt intake ⁸⁸
	Alcohol intake ¹⁴⁹
Hyperglycemia	Hyperglycemia and Diabetes ¹⁰⁰
	High glycemic diet ⁹⁸
Low Oxygen	Hypoxia (Sleep apnea) ¹¹⁶
Ischemia	Myocardial Infarction, Acute Kidney Injury ^{92,101}
Fructose Metabolism by KHK (such as GCK activation)	Western diet-induced steatosis ¹⁴⁸
Heat Stress	Heat Stress-induced kidney injury ⁹⁵
Physical or Emotional Stress	Following major surgery ⁹⁶
Pregnancy	Brain production ⁹⁷
Uric acid	Hyperuricemia ⁶³

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